Mathematical modelling of transport and signalling processes in cells and tissues.

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Mathematical model of a simplest gene regulatory network: canonical Hes1 GRN

Plant hormones: Modelling of Auxin signalling pathway

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Intercellular transport of signalling molecules

The **Cell** (from Latin cella, meaning "small room") is the basic structural, functional, and biological unit of all known living organisms. Cells are the smallest unit of life that can replicate independently, and are often called the "building blocks of life".

Wikipedia





Elodea leaf

Epithelial tissue



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Cell signalling and Gene regulatory networks

Cell signalling: the ability of cells to perceive and correctly respond to their microenvironment is the basis of development, tissue repair, and immunity as well as normal tissue homeostasis.

Errors in cellular information processing are responsible for diseases such as cancer, autoimmunity, abnormal growth in plants. By understanding cell signalling, diseases may be treated effectively.

Signaling molecules interact with a target cell as a ligand to cell surface receptors, and/or by entering into the cell through its membrane or endocytosis for intracellular signaling.

Wikipedia

Gene regulatory networks are at the heart of intercellular signal transduction and control many important cellular functions.

Modelling of Signaling Pathways and Gene Regulatory network



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Modelling of signaling processes

Logical models: the state transition from time t to time t + 1 is defined for each x_i by a Boolean function f

$$x_i(t+1) = f_i(\mathbf{x}(t)), \quad i = 1, ..., N, \quad \mathbf{x} = (x_1, ..., x_N)$$

Differential equations: using the Law of Mass Action

$$\frac{d\mathbf{x}(t)}{dt} = F(t, \mathbf{x}(t))$$

Stochastic models: chemical master equation

$$\frac{\partial P(x,t)}{\partial t} = \sum_{j=1}^{M} \left[a_j(x-\nu_j) P(x-\nu_j,t) - a_j(x) P(x,t) \right]$$

the probability of the system being in a particular state x over dt, a_j probability for a reaction to occur in the interval [t, t + dt).

 Bayesian models is a probabilistic graphical model: to analyse gene expression data.

Gene Regulatory Network

- Gene regulatory network (GRN): collection of DNA segments in a cell which interact with each other to govern the gene expression levels of mRNA and proteins
- Hes1 contributes to heterogeneous differentiation responses of embryonic stem cells (nervous and digestive systems)
- Hes1 enhances the self-renewal and tumourigenicity of stem-like cancer cells in colon cancer
- Hes1 can repress its own expression by directly binding to N-box target sequences in its own promoter

Hes1 gene expression



- T. Kobayashi, R. Kageyama, Genes 2011
 - transcription of Hes1 mRNA by a transcription factor
 - translation of Hes1 protein from Hes1 mRNA
 - decay of Hes1 mRNA and protein

 \rightarrow Hes1 mRNA (m) \rightarrow Hes1 protein (p)

 \rightarrow Hes1 mRNA (*m*) \rightarrow Hes1 protein (*p*)



$$\frac{dm}{dt} = \alpha_m - \mu_m \, m \qquad = F(m, p)$$

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 \rightarrow Hes1 mRNA (*m*) \rightarrow Hes1 protein (*p*)



$$\frac{dm}{dt} = \alpha_m - \mu_m m = F(m, p)$$
$$\frac{dp}{dt} = \alpha_p m - \mu_p p = G(m, p)$$

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 \rightarrow Hes1 mRNA (*m*) \rightarrow Hes1 protein (*p*)

$$\frac{dm}{dt} = \frac{\alpha_m}{1 + [p]^h} - \mu_m m = F(m, p)$$
$$\frac{dp}{dt} = \alpha_p m - \mu_p p = G(m, p)$$

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for some h > 1, $f(p) = \frac{1}{1 + [p]^h}$

 \rightarrow Hes1 mRNA (*m*) \rightarrow Hes1 protein (*p*)



Steady state solutions (m^*, p^*) , i.e. $\frac{dm}{dt} = 0$, $\frac{dp}{dt} = 0$,

$$\begin{split} G(m^*,p^*) &= 0, \qquad m^* = \frac{\mu_p}{\alpha_p} p^*, \\ F(m^*,p^*) &= 0, \qquad p^* + (p^*)^{h+1} = \frac{\alpha_m \alpha_p}{\mu_p \mu_m}, \qquad \text{choose } p^* > 0 \end{split}$$

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Consider small perturbations of the steady state

$$m(t) = m^* + \tilde{m}(t), \qquad p(t) = p^* + \tilde{p}(t), \qquad \text{with} \quad \|\tilde{m}\|, \|\tilde{p}\| \ll 1$$
$$\frac{d\tilde{m}}{dt} = F(m^* + \tilde{m}, p^* + \tilde{p})$$
$$\frac{d\tilde{p}}{dt} = G(m^* + \tilde{m}, p^* + \tilde{p})$$

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$$\frac{d\tilde{m}}{dt} = F(m^*, p^*) + \frac{\partial F(m^*, p^*)}{\partial m} \tilde{m} + \frac{\partial F(m^*, p^*)}{\partial p} \tilde{p} + \dots$$
$$\frac{d\tilde{p}}{dt} = G(m^*, p^*) + \frac{\partial G(m^*, p^*)}{\partial m} \tilde{m} + \frac{\partial G(m^*, p^*)}{\partial p} \tilde{p} + \dots$$

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apply Taylor expansion about (m^*, p^*)

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$$\frac{d\tilde{m}}{dt} = = \qquad \qquad \frac{\partial F(m^{*}, p^{*})}{\partial m} \tilde{m} + \frac{\partial F(m^{*}, p^{*})}{\partial p} \tilde{p} + \dots$$
$$\frac{\partial G(m^{*}, p^{*})}{\partial m} \tilde{m} + \frac{\partial G(m^{*}, p^{*})}{\partial p} \tilde{p} + \dots$$

Linearised system

$$\frac{d}{dt}\begin{pmatrix}\tilde{m}\\\tilde{p}\end{pmatrix} = J(m^*, p^*)\begin{pmatrix}\tilde{m}\\\tilde{p}\end{pmatrix}, \qquad J(m^*, p^*) = \begin{pmatrix}\frac{\partial F(m^*, p^*)}{\partial m} & \frac{\partial F(m^*, p^*)}{\partial p}\\\frac{\partial G(m^*, p^*)}{\partial m} & \frac{\partial G(m^*, p^*)}{\partial p}\end{pmatrix}$$

Small perturbations are solutions of the linear system

$$rac{d}{dt} \begin{pmatrix} ilde{m} \\ ilde{p} \end{pmatrix} = J(m^*,p^*) \begin{pmatrix} ilde{m} \\ ilde{p} \end{pmatrix}$$

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Consider

$$ilde{m}(t) = e^{\lambda t} \hat{m}, \quad ilde{p}(t) = e^{\lambda t} \hat{p}, \qquad \hat{m}, \hat{p} \in \mathbb{R}, \qquad \hat{m} \neq 0, \quad \hat{p} \neq 0$$

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eq0,\quad \hat{p}
eq0$$

Then

$$\begin{bmatrix} J(m^*,p^*)-\lambda I \end{bmatrix} \begin{pmatrix} \hat{m} \\ \hat{p} \end{pmatrix} = 0 \quad \text{and} \quad \det \left(J(m^*,p^*)-\lambda I \right) = 0.$$

where
$$J(m^*, p^*) = \begin{pmatrix} -\mu_p & -\alpha_m \gamma_m \\ \alpha_p & -\mu_p \end{pmatrix},$$

with $\gamma_m = -f'(p^*) = h[p^*]^{h-1}/(1+[p^*]^h)^2$ and $\gamma_m > 0$.

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$$\det \left(J(m^*, p^*) - \lambda I \right) = \lambda^2 + (\mu_m + \mu_p)\lambda + (\alpha_m \alpha_p \gamma_m + \mu_m \mu_p) = 0$$

$$\lambda_{1,2} = \frac{1}{2} \left[-(\mu_m + \mu_p) \pm \sqrt{(\mu_m + \mu_p)^2 - 4(\alpha_m \alpha_p \gamma_m + \mu_m \mu_p)} \right]$$

 $\implies \operatorname{Re}(\lambda_{1,2}) < 0 \implies (m^*, p^*) \text{ is stable.}$

DDE model - Monk (Current Biology 2003)



$$\frac{dm(t)}{dt} = \frac{\alpha_m}{1 + [p(t)]^h} - \mu_m m(t)$$
$$\frac{dp(t)}{dt} = \alpha_p m(t) - \mu_p p(t)$$

no oscillatory behaviour stable steady state

Adding delay produces oscillatory dynamics

$$\frac{dm(t)}{dt} = \frac{\alpha_m}{1 + \left[p(t - \tau_m)\right]^h} - \mu_m m(t), \qquad h > 1$$

$$\frac{dp(t)}{dt} = \alpha_p m(t - \tau_p) - \mu_p p(t)$$

$$m(t) = m_0(t) \quad t \in [-\tau_m, 0], \quad p(t) = p_0(t) \quad t \in [-\tau_p, 0]$$

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DDE model - Monk (Current Biology 2003)



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Small perturbations $m(t) = m^* + \tilde{m}(t)$, $p(t) = p^* + \tilde{p}(t)$:

$$\frac{d\tilde{m}(t)}{dt} = -\alpha_m \gamma_m \tilde{p}(t - \tau_m) - \mu_m \tilde{m}(t)$$
$$\frac{d\tilde{p}(t)}{dt} = \alpha_p \tilde{m}(t - \tau_p) - \mu_p \tilde{p}(t)$$

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where $\gamma_m = -f'(p^*) = h[p^*]^{h-1}/(1+[p^*]^h)^2$, $\gamma_m > 0$.

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Consider $\tilde{m}(t) = e^{\lambda t} \hat{m}$ and $\tilde{p}(t) = e^{\lambda t} \hat{p}$ and obtain

$$\det \begin{pmatrix} -\mu_m - \lambda & -\gamma_m \alpha_m e^{-\lambda \tau_m} \\ \alpha_p e^{-\lambda \tau_p} & -\mu_p - \lambda \end{pmatrix} = 0$$

$$\lambda^{2} + (\mu_{m} + \mu_{p})\lambda + \mu_{m}\mu_{p} + \gamma_{m}\alpha_{m}\alpha_{p}e^{-\lambda\tau} = 0, \qquad \tau = \tau_{m} + \tau_{p}$$

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Consider $\tilde{m}(t) = e^{\lambda t} \hat{m}$ and $\tilde{p}(t) = e^{\lambda t} \hat{p}$ and obtain

 \Leftarrow

$$\det \begin{pmatrix} -\mu_m - \lambda & -\gamma_m \alpha_m e^{-\lambda \tau_m} \\ \alpha_p e^{-\lambda \tau_p} & -\mu_p - \lambda \end{pmatrix} = 0$$

$$\Rightarrow \qquad \lambda^2 + (\mu_m + \mu_p)\lambda + \mu_m \mu_p + \gamma_m \alpha_m \alpha_p e^{-\lambda \tau} = 0, \qquad \tau = \tau_m + \tau_p$$

For $\gamma_m \alpha_m \alpha_p > \mu_m \mu_p$ there exist $\omega_0 > 0$ such that $\lambda = \pm i\omega_0$ single pair of pure imaginary eigenvalues for $\tau_j = \frac{1}{\omega_0} \Big[\sin^{-1} \Big(\frac{\mu_m + \mu_p}{\alpha_p \alpha_m \gamma_m} \omega_0 \Big) + 2\pi j \Big], \quad j = 0, 1, 2, \dots$

Small perturbations $m(t) = m^* + \tilde{m}(t)$, $p(t) = p^* + \tilde{p}(t)$:

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$$\det \begin{pmatrix} -\mu_m - \lambda & -\gamma_m \alpha_m e^{-\lambda \tau_m} \\ \alpha_p e^{-\lambda \tau_p} & -\mu_p - \lambda \end{pmatrix} = 0$$

$$\iff \lambda^2 + (\mu_m + \mu_p)\lambda + \mu_m \mu_p + \gamma_m \alpha_m \alpha_p e^{-\lambda \tau} = 0, \qquad \tau = \tau_m + \tau_p$$

For $\gamma_m \alpha_m \alpha_p > \mu_m \mu_p$ there exist $\omega_0 > 0$ such that $\lambda = \pm i \omega_0$ single pair of pure imaginary eigenvalues

$$\tilde{m}(t) = e^{\lambda t} \hat{m}, \quad \tilde{p}(t) = e^{\lambda t} \hat{p}, \quad e^{i\omega_0 t} = \cos(\omega_0 t) + i\sin(\omega_0 t)$$

Stability analysis of delay differential equations $\lambda^{2} + (\mu_{m} + \mu_{p})\lambda + \mu_{m}\mu_{p} + \gamma_{m}\alpha_{m}\alpha_{p}e^{-\lambda\tau} = 0$

Considering the eigenvalues $\lambda = \lambda(\tau)$ as functions of τ we have

 $\frac{d}{d\tau}(\operatorname{Re}\lambda)(\tau_j)>0, \quad j=0,1,2,\ldots \Longrightarrow (m^*,p^*) \text{ unstable for all } \tau\geq \tau_0$

Stability analysis of delay differential equations $\lambda^2 + (\mu_m + \mu_p)\lambda + \mu_m\mu_p + \gamma_m\alpha_m\alpha_p e^{-\lambda\tau} = 0$

Considering the eigenvalues $\lambda = \lambda(\tau)$ as functions of τ we have

 $\frac{d}{d\tau}(\operatorname{Re}\lambda)(\tau_j) > 0, \quad j = 0, 1, 2, \ldots \implies (m^*, p^*) \text{ unstable for all } \tau \geq \tau_0$

Theorem (Hopf Bifurcation)

- (F, G) is continuously differentiable in (m, p) and τ .
- $(m^*(\tau), p^*(\tau))$ isolate stationary solution for $\tau \ge 0$
- ▶ for $\tau = \tau_j$ there exists a simple pair of pure imaginary eigenvalues $\lambda = \pm i\omega_0$ with $\omega_0 \neq 0$
- ▶ for all $n \in \mathbb{Z} \setminus \{1, -1\}$, $\pm i \, n \, \omega_0$ are not eigenvalue
- near τ_j we have the eigenvalues λ = r(τ) ± iω(τ), with r and ω are continuous and r(τ_j) = 0, ω(τ_j) = ω₀

• transversality condition:
$$\frac{d}{d\tau}(\text{Re}\lambda)(\tau_j) = \frac{dr}{d\tau}(\tau_j) \neq 0$$

Then the system has periodic solutions in a neighborhood of τ_j , bifurcating from the stationary solution.

Hes1 gene expression oscillation



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T. Kobayashi, R. Kageyama, Genes 2011

Interaction between cell nucleus and cell cytoplasm: transcription (mRNA production) in nucleus and translation (protein production) in cytoplasm

Model Schematic



$$\begin{aligned} \frac{\partial m}{\partial t} &= + \frac{\alpha_m}{1 + p^h} - \mu m & \text{in } (0, T) \times (0, 1) \\ \frac{\partial p}{\partial t} &= + \alpha_p m & - \mu p & \text{in } (0, T) \times (0, 1), \\ m(0) &= m_0, \quad p(0) &= p_0 & \text{in } (0, 1), \end{aligned}$$



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 x_{M} - position of the centre of the gene site, (1, 1)- cell cytoplasm

$$\begin{aligned} \frac{\partial m}{\partial t} &= + \frac{\alpha_m}{1 + p^h} \delta^{\varepsilon}_{x_M}(x) - \mu m & \text{in } (0, T) \times (0, 1) \\ \frac{\partial p}{\partial t} &= + \alpha_p m g(x) - \mu p & \text{in } (0, T) \times (0, 1), \\ m(0) &= m_0, \quad p(0) = p_0 & \text{in } (0, 1), \end{aligned}$$

$$\delta_{x_M}^{\varepsilon} - \text{Dirac sequence}, \qquad g(x) = \begin{cases} 0, & \text{if } x < l \\ \\ 1, & \text{if } x \ge l \end{cases}$$

 x_{M} - position of the centre of the gene site, (1,1)- cell cytoplasm

$$\begin{aligned} \frac{\partial m}{\partial t} &= D \frac{\partial^2 m}{\partial x^2} + \frac{\alpha_m}{1 + p^h} \, \delta^{\varepsilon}_{x_M}(x) - \mu \, m & \text{in } (0, T) \times (0, 1) \\ \frac{\partial p}{\partial t} &= D \frac{\partial^2 p}{\partial x^2} + \alpha_p \, m \, g(x) - \mu \, p & \text{in } (0, T) \times (0, 1), \\ m(0) &= m_0, \quad p(0) = p_0 & \text{in } (0, 1), \end{aligned}$$

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$$\frac{\partial p}{\partial t} = D \frac{\partial^2 p}{\partial x^2} + \alpha_p m g(x) - \mu p \qquad \text{in } (0, T) \times (0, 1),$$

$$m(0) = m_0, \quad p(0) = p_0 \qquad \text{in } (0, 1),$$

$$m_x(t, 0) = m_x(t, 1) = p_x(t, 0) = p_x(t, 1) = 0 \qquad \text{in } (0, T)$$

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 x_{M} - position of the centre of the gene site, (1,1)- cell cytoplasm









Hopf Bifurcation for GRN Eigenvalue problem

$$\begin{split} \lambda \bar{m}^{\varepsilon} &= D \bar{m}_{xx}^{\varepsilon} - \mu \bar{m}^{\varepsilon} + \alpha_m f'(p_{\varepsilon}^*(x,D)) \, \delta_{x_M}^{\varepsilon} \, \bar{p}^{\varepsilon} & \text{ in } (0,1) \\ \lambda \bar{p}^{\varepsilon} &= D \bar{p}_{xx}^{\varepsilon} - \mu \bar{p}^{\varepsilon} + \alpha_p g(x) \bar{m}^{\varepsilon} & \text{ in } (0,1) \\ + \text{ zero-flux b.c.} \end{split}$$

For $\lambda \in \mathbb{C}$ such that $\mathcal{R}e(\lambda) > -\mu$ or $\mathcal{I}m(\lambda) \neq 0$:

$$\bar{m}^{\varepsilon}(x) = \alpha_{m}(\lambda I - \mathcal{A}_{0})^{-1} \left(f'(p_{\varepsilon}^{*}(x, D)) \bar{p}^{\varepsilon}(x) \delta_{x_{M}}^{\varepsilon}(x) \right)$$
$$\lambda \bar{p}^{\varepsilon} = D \frac{d^{2} \bar{p}^{\varepsilon}}{dx^{2}} + \alpha_{p} \alpha_{m} g(x) (\lambda I - \mathcal{A}_{0})^{-1} \left(f'(p_{\varepsilon}^{*}) \bar{p}^{\varepsilon} \delta_{x_{M}}^{\varepsilon} \right) - \mu \bar{p}^{\varepsilon}$$
$$\bar{p}_{x}^{\varepsilon}(0) = \bar{p}_{x}^{\varepsilon}(1) = 0.$$

where $\mathcal{A}_0 = (D \frac{d^2}{dx^2} - \mu).$

Theorem For $\varepsilon > 0$ small there exist two critical values $D_{1,\varepsilon}^c$ and $D_{2,\varepsilon}^c$ for which a Hopf bifurcation occurs in the canonical Hes1 GRN model.

A bit more complicated network: Auxin signalling

- Auxins belong to the most important plant hormones and play a central role in growth and development regulation. (direction of growth, changes in shoot and root branching and changes in vascular differentiation).
- Transcription factor ARF (auxin response factor) activates Aux/IAA gene and the transcription of Aux/IAA mRNA
- The accumulation of Aux/IAA protein stimulates the formation of ARF:Aux/IAA protein complexes, which repress Aux/IAA genes and inhibits the production of Aux/IAA mRNA.
- ► When the levels of Aux/IAA proteins fall, the concentration of free ARFs increases, enhancing Aux/IAA transcription and translation.
- Auxin functions by mediating the activation of the Aux/IAA family of genes by auxin mediating turnover of Aux/IAA proteins (ubiquitination)
- The instability of Aux/IAA proteins is required for normal auxin signalling.

Auxin signalling



Vernoux et al. Molecular Systems Biol. 2011

Aux:TIR1 (r_b) targets Aux/IAA protein (p) by forming Aux:TIR1:IAA (p_b) whose dissociation results in the ubiquitin-tagged protein Aux/IAA* (p_u)

$$\begin{array}{l} \operatorname{Aux}:\operatorname{TIR1}+\operatorname{IAA} \stackrel{\beta_r}{\rightleftharpoons} \operatorname{Aux}:\operatorname{TIR1}:\operatorname{IAA} \stackrel{\beta_{pb}}{\to} \operatorname{Aux}:\operatorname{TIR1}+\operatorname{IAA}^* \\ \gamma_r \end{array}$$

Auxin synthesised and degrade at the constant rates $lpha_A$ and μ_A

$$\begin{array}{ccc} \alpha_A & \mu_A & \mu_u \\ \rightarrow & \mathrm{Aux} \rightarrow & \emptyset, & \mathrm{IAA}^* \rightarrow & \emptyset \end{array}$$

$$ARF + ARF \rightleftharpoons ARF_{2}, \qquad IAA + ARF \rightleftharpoons ARF : IAA$$
$$\gamma_{R} \qquad \gamma_{P}$$

Aux:TIR1 (r_b) targets Aux/IAA protein (p) by forming Aux:TIR1:IAA (p_b) whose dissociation results in the ubiquitin-tagged protein Aux/IAA* (p_u)

$$\begin{array}{l} \operatorname{Aux}:\operatorname{TIR1}+\operatorname{IAA} \stackrel{\beta_r}{\rightleftharpoons} \operatorname{Aux}:\operatorname{TIR1}:\operatorname{IAA} \stackrel{\beta_{pb}}{\rightarrow} \operatorname{Aux}:\operatorname{TIR1}+\operatorname{IAA}^* \\ \gamma_r \end{array}$$

Auxin synthesised and degrade at the constant rates $lpha_A$ and μ_A

$$\begin{array}{ccc} \alpha_A & \mu_A & \mu_u \\ \rightarrow & \mathrm{Aux} \rightarrow & \emptyset, & \mathrm{IAA}^* \rightarrow & \emptyset \end{array}$$

$$ARF + ARF \rightleftharpoons ARF_{2}, \qquad IAA + ARF \rightleftharpoons ARF : IAA$$
$$\gamma_{R} \qquad \gamma_{P}$$

Aux:TIR1 (r_b) targets Aux/IAA protein (p) by forming Aux:TIR1:IAA (p_b) whose dissociation results in the ubiquitin-tagged protein Aux/IAA* (p_u)

$$\begin{array}{l} \operatorname{Aux}:\operatorname{TIR1}+\operatorname{IAA} \stackrel{\beta_{r}}{\rightleftharpoons} \operatorname{Aux}:\operatorname{TIR1}:\operatorname{IAA} \stackrel{\beta_{pb}}{\rightarrow} \operatorname{Aux}:\operatorname{TIR1}+\operatorname{IAA}^{*} \\ \gamma_{r} \end{array}$$

Auxin synthesised and degrade at the constant rates α_A and μ_A

$$\stackrel{\alpha_{\mathcal{A}}}{\to} \operatorname{Aux} \stackrel{\mu_{\mathcal{A}}}{\to} \emptyset, \qquad \operatorname{IAA}^* \stackrel{\mu_{\mathcal{U}}}{\to} \emptyset$$

$$ARF + ARF \rightleftharpoons ARF_{2}, \qquad IAA + ARF \rightleftharpoons ARF : IAA$$
$$\gamma_{R} \qquad \gamma_{P}$$

$$\begin{array}{l} \operatorname{Aux} + \operatorname{TIR1} \begin{array}{c} \beta_{\mathsf{A}} \\ \rightleftharpoons \end{array} \operatorname{Aux} : \operatorname{TIR1} \\ \gamma_{\mathsf{A}} \end{array}$$

Aux:TIR1 (r_b) targets Aux/IAA protein (p) by forming Aux:TIR1:IAA (p_b) whose dissociation results in the ubiquitin-tagged protein Aux/IAA* (p_u)

$$\begin{array}{l} \operatorname{Aux}:\operatorname{TIR1}+\operatorname{IAA} \stackrel{\beta_r}{\rightleftharpoons} \operatorname{Aux}:\operatorname{TIR1}:\operatorname{IAA} \stackrel{\beta_{pb}}{\to} \operatorname{Aux}:\operatorname{TIR1}+\operatorname{IAA}^* \\ \gamma_r \end{array}$$

Auxin synthesised and degrade at the constant rates α_A and μ_A

$$\stackrel{\alpha_{\mathcal{A}}}{\to} \operatorname{Aux} \stackrel{\mu_{\mathcal{A}}}{\to} \emptyset, \qquad \operatorname{IAA}^* \stackrel{\mu_{\mathcal{U}}}{\to} \emptyset$$

$$ARF + ARF \rightleftharpoons^{\beta_{R}}_{P} ARF_{2}, \qquad IAA + ARF \rightleftharpoons^{\beta_{p}}_{P} ARF : IAA$$
$$\gamma_{R} \qquad \gamma_{P}$$

Auxin Signalling Network:Law of Mass Action $\frac{dA}{dt} = \alpha_A - \mu_A A - \beta_A A r_f + \gamma_A r_b$ $\stackrel{A}{\operatorname{Aux}} + \operatorname{TIR1}^{r_f} \rightleftharpoons \operatorname{Aux}^{r_b} \operatorname{TIR1}^{r_b}$

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Auxin Signalling Network: Law of Mass Action

$$\frac{dA}{dt} = \alpha_A - \mu_A A - \beta_A A r_f + \gamma_A r_b$$

$$\frac{dr_f}{dt} = -\beta_A A r_f + \gamma_A r_b$$

$$\frac{dr_f}{dt} = -\beta_A A r_f - \gamma_A r_b - \beta_r r_b p + (\gamma_r + \beta_{pb}) p_b$$

$$\frac{dp_b}{dt} = \beta_r r_b p - (\gamma_r + \beta_{pb}) p_b$$

$$Aux + TIR1 \stackrel{\beta_A}{\Rightarrow} Aux : TIR1 + IAA$$

$$\frac{\beta_r}{\beta_{pb}} \stackrel{r_b}{\Rightarrow} Aux : TIR1 : IAA$$

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Auxin Signalling Network:Law of Mass Action
$$\frac{dA}{dt} = \alpha_A - \mu_A A - \beta_A A r_f + \gamma_A r_b$$
 $\stackrel{A}{Aux} + TIR1 \stackrel{\beta}{=} Aux : TIR1$ $\frac{dr_f}{dt} = -\beta_A A r_f + \gamma_A r_b$ $\stackrel{A}{Aux} + TIR1 \stackrel{\beta}{=} Aux : TIR1$ $\frac{dr_f}{dt} = -\beta_A A r_f + \gamma_A r_b$ $\stackrel{A}{aux} + TIR1 \stackrel{\beta}{=} Aux : TIR1 + IAA$ $\frac{dr_b}{dt} = \beta_A A r_f - \gamma_A r_b - \beta_r r_b p + (\gamma_r + \beta_{pb})p_b$ $\stackrel{\beta_r}{=} Aux : TIR1 + IAA$ $\frac{dp_b}{dt} = \beta_r r_b p - (\gamma_r + \beta_{pb})p_b$ $\stackrel{mRNA (m) production by ARF, ARF: inhibition by ARF: IAA (R_p)$

$$\frac{dp}{dt} = \alpha_p m - \beta_r r_b p + \gamma_r p_b - \beta_p p R + \gamma_p R_p$$

2 inhibition by ARF: IAA (R_p)

Auxin Signalling Network: Law of Mass Action $\frac{dA}{dt} = \alpha_A - \mu_A A - \beta_A A r_f + \gamma_A r_b$ $\frac{dt}{dt} = -\beta_A Ar_f + \gamma_A r_b$ $\frac{dr_b}{dt} = \beta_A A r_f - \gamma_A r_b - \beta_r r_b p + (\gamma_r + \beta_{pb})p_b$ $\frac{dp_b}{dt} = \beta_r r_b p - (\gamma_r + \beta_{pb})p_b$ $\frac{dm}{dt} = f(R, R_p, R_2) - \mu_m m$ mRNA (m) production by ARF, ARF₂ inhibition by ARF: IAA (R_p) $\frac{dp}{dt} = \alpha_{p}m - \beta_{r}r_{b}p + \gamma_{r}p_{b} - \beta_{p}pR + \gamma_{p}R_{p}$ $\frac{dR}{dt} = -2\beta_{R}R^{2} + 2\gamma_{R}R_{2} - \beta_{p}pR + \gamma_{p}R_{p}$ $\frac{dR_{p}}{dt} = \beta_{p}pR - \gamma_{p}R_{p}$ $\begin{cases} \text{ARF} + \text{ARF} \rightleftharpoons \beta_{R} \\ \gamma_{R} \\ \beta_{R} \\ \beta_{$

$$\frac{dt}{dt} = \beta_p \, p \, R - \gamma_p \, R_p$$

$$\frac{dR_2}{dt} = \beta_R \, R^2 - \gamma_R \, R_2$$

$$\int IAA + ARF \rightleftharpoons ARF : IAA$$

$$\gamma_p$$

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Auxin Signalling Network: Law of Mass Action $\frac{dA}{dt} = \alpha_A - \mu_A A - \beta_A A r_f + \gamma_A r_b$ $\frac{dr_f}{dt} = -\beta_A A r_f + \gamma_A r_b$ $dr_c dr_b dp_b$

$$\frac{dr_b}{dt} = \beta_A A r_f - \gamma_A r_b - \beta_r r_b p + (\gamma_r + \beta_{pb}) p_b$$
$$\frac{dp_b}{dt} = \beta_r r_b p - (\gamma_r + \beta_{rb}) p_b$$

$$\frac{dr_f}{dt} + \frac{dr_b}{dt} + \frac{dp_b}{dt} = 0$$

 $\frac{1}{dt} = \beta_r r_b p - (\gamma_r + \beta_{pb}) p_b$ $\frac{dm}{dt} = f(R, R_p, R_2) - \mu_m m$

mRNA (m) production by ARF, ARF_2 inhibition by ARF:IAA (R_p)

$$\frac{dp}{dt} = \alpha_{p}m - \beta_{r}r_{b}p + \gamma_{r}p_{b} - \beta_{p}pR + \gamma_{p}R_{p}$$

$$\frac{dR}{dt} = -2\beta_{R}R^{2} + 2\gamma_{R}R_{2} - \beta_{p}pR + \gamma_{p}R_{p}$$

$$\frac{dR_{p}}{dt} = \beta_{p}pR - \gamma_{p}R_{p}$$

$$\frac{dR_{2}}{dt} = \beta_{R}R^{2} - \gamma_{R}R_{2}$$

$$\begin{cases}
\frac{dR}{dt} + \frac{dR_{p}}{dt} + 2\frac{dR_{2}}{dt} = 0$$

where $f(R, R_p, R_2) = (\alpha_m R + 2\beta_m R_2 + \gamma_m R^2)/(1 + \kappa_m R_p + \zeta_m p R) = \gamma_Q P$

Auxin Signalling Network

Assume that total concentrations of TIR1 (r_f) and ARF (T) are constant:

$$p_{b} + r_{f} + r_{b} = r_{f,int}, \quad R + R_{p} + 2R_{2} = R_{int}.$$

$$\frac{dA}{dt} = -\beta_{A} A r_{f} + \gamma_{A} r_{b} + \alpha_{A} - \mu_{A} A,$$

$$\frac{dr_{f}}{dt} = -\beta_{A} A r_{f} + \gamma_{A} r_{b}$$

$$\frac{dr_{b}}{dt} = \beta_{A} A r_{f} - \gamma_{A} r_{b} - \beta_{r} r_{b} p + (\gamma_{r} + \beta_{pb})(r_{f,int} - r_{b} - r_{f})$$

$$\frac{dm}{dt} = F(R, R_{p}, R_{int}) - \mu_{m} m,$$

$$\frac{dp}{dt} = \alpha_{p} m - \beta_{r} r_{b} p + \gamma_{r}(r_{f,int} - r_{b} - r_{f}) - \beta_{p} p R + \gamma_{p} R_{p},$$

$$\frac{dR}{dt} = -2\beta_{R} R^{2} + \gamma_{R}(R_{int} - R_{p} - R) - \beta_{p} p R + \gamma_{p} R_{p}$$

$$\frac{dR_{p}}{dt} = \beta_{p} p R - \gamma_{p} R_{p}$$

$$\frac{dp_{u}}{dt} = \beta_{pb} (r_{f,int} - r_{b} - r_{f}) - \mu_{u} p_{u},$$

$$F(R, R_{p}, R_{int}) = (\alpha_{m} R + \beta_{m}(R_{int} - R_{p} - R) + \gamma_{m} R_{p}^{2})/(1 + \kappa_{m} R_{p} + \zeta_{m} p R) \rightarrow \infty$$

- Non-dimesionalization: rates of formation and dissociation of ARF₂ and the effective rate of Aux/IAA translation (translation rate/ production of ubiquitin-tagged protein) are key parameters
- Long time behaviour: exists a single steady-state.
- Stability analysis: three parameter regimes
 - mono-stable: steady-state is stable or it is unstable and there exists a stable limit-cycle (slow formation and dissociation of ARF₂ and large effective rate of translation)
 - bi-stable: steady-state is stable and there exist a stable and an unstable limit-cycles



Intercellular transport of signalling molecules



http://www.quia.com/jg/1225839list.html

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Signalling molecules interact with a target cell

- as a ligand to cell surface receptors
- and/or by entering into the cell through its membrane or endocytosis for intracellular signaling

Intercellular transport of signalling molecules



J. Downward, Nature 2001

- Consider signalling molecules (ligands) / in the intercellular space and receptors on the cell membrane
- Free and bound receptors r_f and r_b
- Cells produce new receptors r_f and signalling molecules (ligands)
- Ligands / diffuse in the intercellular space and bind to the receptors on the membrane
- Bound receptors r_b dissociate back to free receptors and ligands
- All the considered molecules undergo natural decay

 Diffusion, production and decay of signalling molecules (ligands) in the intercellular space

$$\frac{\partial}{\partial t}I = \nabla \cdot (D\nabla I) + p_I(I) - \mu_I I$$



Binding on the cell surfaces

$$D \nabla l \cdot \nu = -b l r_f + d r_b$$
 on Γ

- / density of ligands
- μ_I rate of decay of ligands,
- p_l production of ligands,
- D diffusion coefficient.

 Diffusion, production and decay of signalling molecules (ligands) in the intercellular space

$$\frac{\partial}{\partial t}I = \nabla \cdot (D\nabla I) + p_I(I) - \mu_I I$$

 Interaction between a signalling molecule L and a free receptor R_f results into a bound receptor R_b

$$L+R_f \stackrel{b}{\rightleftharpoons} R_k$$



binding process is governed by the Law of Mass Action

Binding on the cell surfaces

$$D \nabla l \cdot \nu = -b l r_f + d r_b$$
 on Γ

- / density of ligands
- μ_I rate of decay of ligands,
- p_l production of ligands, D
- diffusion coefficient.

 Diffusion, production and decay of signalling molecules (ligands) in the intercellular space

$$\frac{\partial}{\partial t}I = \nabla \cdot (D\nabla I) + p_I(I) - \mu_I I$$

 Interaction between a signalling molecule L and a free receptor R_f results into a bound receptor R_b

$$L+R_f \stackrel{b}{\rightleftharpoons} R_b$$



binding process is governed by the Law of Mass Action

Binding on the cell surfaces

$$D \nabla l \cdot \nu = -b l r_f + d r_b$$
 on Γ

- / density of ligands μ_l rate of decay of ligands,
- p_l production of ligands, D diffusion coefficient.

Mathematical modelling of signalling processes

Reaction equations for the receptors on the cell surface Γ

$$\frac{\partial}{\partial t}r_{f} = -br_{f}l + dr_{b} - \mu_{f}r_{f} \quad \text{on } \Gamma, t > 0$$

$$\frac{\partial}{\partial t}r_{b} = br_{f}l - dr_{b} - \mu_{b}r_{b} \quad \text{on } \Gamma, t > 0$$

with initial conditions

$$r_f(0, x) = r_{f,int}(x)$$
 on Γ
 $r_b(0, x) = r_{b,int}(x)$ on Γ

 $\begin{array}{ll} r_f, & r_b & \text{density of free and bound receptors} \\ \mu_f, & \mu_b & \text{rates of decay of free and bound receptors} \\ d & \text{rate of dissociation of bound receptors} \\ b & \text{rate of binding of ligands and free receptors} \end{array}$



Cell to cell transport of plant hormones Polar transport of auxin

- Auxin (IAA) is mostly produced in the plant shoot and is transported polar from cell to cell through the shoot and stem towards the roots.
- Plant hormones (signalling molecules) regulate plant growth, determine the formation of flowers, stems, leaves, the shedding of leaves, and the development and ripening of fruit
- Aim: To determine the distribution and the transport velocity of the auxin



Auxin (IAA) is a weak acid which dissociates into ions

IAAH
$$\rightleftharpoons^{r_d}_{r_r}$$
 IAA⁻ + H⁺

 r_d and r_r are dissociation and recombination rates.

- IAAH predominates in cell wall due to acidic pH
- ► IAA⁻ predominates in cytoplasm due to neutral pH
- IAAH is uncharged and can diffuse through membrane
- membrane is impermeable to IAA⁻ due to charge
- influx protein AUX1 transports IAA⁻ into cytoplasm
- efflux PIN proteins transport IAA⁻ out of cell

Due to the negative charge, the electric potential differences across the plasma membrane and tonoplast produce an additional flux of IAA⁻. (membrane potential acts on changed IAA⁻: ca. -120 mV between cell wall and cytoplasm and 50 mV between cytoplasm and vacuole).





$$\begin{array}{rcl} \mathrm{IAAH} & \stackrel{r_d}{\rightleftharpoons} & \mathrm{IAA}^- & + & \mathrm{H}^+ \ , \\ & & r_r \end{array}$$

- u the concentrations of the auxin ions IAA⁻
- ▶ *v* the concentrations of the protonated auxin IAAH

- The mobility of the ions is given by the permeability P
- φ is the electric field, assume φ cell is independent of u (concentration of other ions is much larger as of u).
- $P \phi u$ is the electric flux.
- Initial and boundary conditions

$$\begin{array}{rcl} \mathrm{IAAH} & \stackrel{r_d}{\rightleftharpoons} & \mathrm{IAA}^- & + & \mathrm{H}^+ \ , \\ & & r_r \end{array}$$

- ▶ *u* the concentrations of the auxin ions IAA⁻
- ▶ *v* the concentrations of the protonated auxin IAAH

$$\partial_t u = r_d v - r_r u$$

 $\partial_t v = -r_d v + r_r u$

- The mobility of the ions is given by the permeability P
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- u the concentrations of the auxin ions IAA⁻
- \triangleright v the concentrations of the protonated auxin IAAH

$$\partial_t u = \nabla \cdot (D_u \nabla u) + r_d v - r_r u$$

$$\partial_t v = \nabla \cdot (D_v \nabla v) - r_d v + r_r u$$

- The mobility of the ions is given by the permeability P
- φ is the electric field, assume φ cell is independent of u (concentration of other ions is much larger as of u).
- $P \phi u$ is the electric flux.
- Initial and boundary conditions

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- \triangleright v the concentrations of the protonated auxin IAAH

$$\partial_t u = \nabla \cdot (D_u \nabla u) - \nabla \cdot (P \phi u) + r_d v - r_r u$$

$$\partial_t v = \nabla \cdot (D_v \nabla v) - r_d v + r_r u$$

- The mobility of the ions is given by the permeability P
- φ is the electric field, assume φ cell is independent of u (concentration of other ions is much larger as of u).
- $P \phi u$ is the electric flux.
- Initial and boundary conditions

Model parameters and scaling

• $D \nabla u_0 \approx 16 u_0$ in wall $\approx 240 u_0$ in cytoplasm $\approx 10^{-9} u_0$ in plasma membranes • $P \phi u_0 \approx 0.3 u_0 \sim 30 u_0$ in plasma membrane. • Reaction rates: $r_d \approx 5 \cdot 10^9 h^{-1}$ and $r_r \approx 5 \cdot 10^6 \sim 5 \cdot 10^8 h^{-1}$.



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Time scale in the model is the characteristic reaction time the flux terms have to be scaled by $\varepsilon = 10^{-3}$.

$$\begin{aligned} \partial_t u &= \varepsilon \nabla \cdot (D_u \nabla u) - \varepsilon \nabla \cdot (P \phi u) + R_d v - R_r u, \\ \partial_t v &= \varepsilon \operatorname{div} (D_v \nabla v) & - R_d v + R_r u \end{aligned}$$

where $R_d = \varepsilon r_d$, $R_r = \varepsilon r_r$

Microscopic structure

	0.0016			in cell wall, in plasma membrane,			1	0.0016		in cell wall,	
	$3.6\cdot10^{-12}$		12					$2 \cdot 10^{-}$	7	in plasma membrar	ıe,
$D_u = \langle$	0.	0.024		in cytoplasm,		$D_v =$		0.024		in cytoplasm,	
	3.	$3.6 \cdot 10^{-12}$ 0.024		in tonoplast,				$2 \cdot 10^{-}$	7	in tonoplast,	
	0.			in vacuole				0.024		in vacuole	
							_				
	1	0	in cell wall,			1	0		in (cell wall,	
		0.1 in p		lasma membrane,			120	$\cdot 10^3$	in	in plasma membrane,	
P =	= {	0	in c	in cytoplasm,		= {	0	in		cytoplasm,	
		0.2	in t	onoplast,			50 ·	10 ³	in tonoplast,		
		0	in v	acuole			0		in '	vacuole	





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Homogenization



- The aim of homogenization is to give the macroscopic properties by taking the properties of the microscopic structure into account.
- To link macroscopic parameters with microscopic properties of the system.

 Derivation of macroscopic models simplifies numerical simulation.

Homogenization



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- To link macroscopic parameters with microscopic properties of the system.
- Derivation of macroscopic models simplifies numerical simulation.

Solve cell problems

and calculate effective coefficients using physiological data



- $Y = (0, 20 \mu m) \times (0, 100 \mu m)$
- Cell wall and cytoplasm are $1 \mu m$
- ► AUX1: $\{y \in Y | y_2 = 1, 1 \le y_1 \le 19\}\mu m$ with permeability 0.2cm h⁻¹
- ▶ PIN: $\{y \in Y | y_2 = 99, 1 \le y_1 \le 19\}\mu m$ with permeability 0.1 cmh⁻¹.
- Permeability of tonoplast transport protein: 0.2 cmh⁻¹
- ► Scalar diffusivity: cytoplasm and vacuole: $D_w = 0.024 \text{ cm}^2 \text{s}^{-1}$; cell wall: $D_w/15$; membrane: $2 \cdot 10^{-7} \text{ cm}^2 \text{s}^{-1}$ for IAAH and $3.6 \cdot 10^{-12} \text{cm}^2 \text{s}^{-1}$ for IAA⁻.
- ▶ Cell wall pH = 5.8, vacuole pH = 5.7 cytoplasm $pH = 7.6 \Rightarrow R_r^{\varepsilon} = R_d \ 10^{pKa-pH^{\varepsilon}}$, where $R_d = 5 \cdot 10^9 \ h^{-1} = \text{const.}$
- The constant approximation for the electric field: membrane potential: -120mV; tonoplast potential: 50mV

Numerical results: Auxin transport velocity



Solutions of the cell problems: w_x (diffusion), Z_{uu} (transport).

$$\frac{\mathcal{A}_{u}}{D_{w}} = \left(\begin{array}{cc} 1.37 & \mathcal{O}(10^{-12}) \\ \mathcal{O}(10^{-12}) & 6.7 \end{array} \right) \cdot 10^{-3}, \ \frac{\mathcal{A}_{v}}{D_{w}} = \left(\begin{array}{cc} 2.85 & \mathcal{O}(10^{-11}) \\ \mathcal{O}(10^{-11}) & 21.6 \end{array} \right) \cdot 10^{-3} \,.$$

$$\begin{split} \mathcal{V}_{uu} &= \left(\begin{array}{c} \mathcal{O}(10^{-4}) \\ 0.638 \end{array} \right) \ cm \ h^{-1} \quad , \quad \mathcal{V}_{uv} = \left(\begin{array}{c} \mathcal{O}(10^{-11}) \\ \mathcal{O}(10^{-7}) \end{array} \right) \ cm \ h^{-1} \, , \\ \mathcal{V}_{vu} &= \left(\begin{array}{c} \mathcal{O}(10^{-11}) \\ \mathcal{O}(10^{-7}) \end{array} \right) \ cm \ h^{-1} \quad , \quad \mathcal{V}_{vv} = \left(\begin{array}{c} \mathcal{O}(10^{-8}) \\ \mathcal{O}(10^{-4}) \end{array} \right) \ cm \ h^{-1} \, . \end{split}$$

Value $\mathcal{V}_{uu} \approx 0.6 \text{ cm h}^{-1}$ near published experimental value. (measurements of pulses of radioactivelly labeled auxin: $1.2 \approx 1.5 \text{ cmh}^{-1}$).

Conclusion

- Different modelling approaches can explain oscillatory behaviour of gene regulatory networks with negative feedback
- Spatial models on the scale of a single cell are considered to describe the intercellular transport of signalling molecules in a plant tissue

Thank you very much for your attention

